

Agromorphological Characterization of Amaranth Accessions

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Abstract

A sound fundamental knowledge of the agro morphological characteristics of amaranth accessions is crucial for promoting their use in food. This review aims to assess the genetic diversity within the collection to understand the range of genetic variation. Thirty-two agromorphological parameters of Ten amaranth accessions were analyzed in Samanko conditions. Wide variability was present concerning leaf pigmentation (PigF), flower density index (IDF), seed color (CH), and Branching index (IR). Statistical analysis showed significant differences among accessions for morphological characters such as number of plants, leaf width, stem height, terminal inflorescence length of stem, axillary inflorescence length, 50% flowering and 1000 grain weight. It appears that the Axillary Inflorescence (LIA) and the Length of the Terminal Inflorescence are correlated ($r = 0.82$). Length of the inflorescence terminal of the stem (LITT) and the Length of the Terminal Basal Branches (LITBT) are associated ($r = 0.75$). Stem height presents a robust correlation ($r = 0.447$) with 50% flowering date. The length of the inflorescence Axillary was also highly correlated with the length of the inflorescence terminal of the stem ($r = 0.904^*$). Ascending hierarchical classification revealed three distinct classes: C1: Madira 1, Madira 2, A2004, A2002; C2: TP5-sel, N'gourouma, Akeri, AC-NL; C3: AHTI, Akeri. "Akeri." "TP5-sel," and "N'gourouma" are associated with the weight of 1000 grains, the length of the terminal branches, and the shape of the terminal inflorescence. "Poly" and "AHTI" are quite similar and are associated with the parameters of the length of basal branches and length of a terminal inflorescence of the stem "Madira 1"; "Madira 2", "A2004", "A2002" on the correlation circle indicates that the height of the plants and the 50% flowering date. These results indicate a high possibility of genetic diversity among the amaranth accessions within the collection. The data can be exploited in future breeding programs to improve the species.

Keywords

Accession, Amaranth, Morphology, Characterization

1. Introduction

African leafy vegetables contribute greatly to achieving food and nutrition security especially in times of famine and natural disasters [1]. According to the work of [2], if people consumed leafy vegetables in their daily diet, it could improve their nutritional status, as these vegetables have high levels of micronutrients. Indigenous leafy vegetables are essential for food and nutrition security. Several studies have shown that African leafy vegetables offer many benefits, high micronutrient content, and beneficial medicinal properties [3] [4]. In addition, these vegetables have interesting agronomic performances, such as a short growth period (some crops are ready to harvest within 3 to 4 weeks after planting) and low input requirements [5]. Amaranth is recognized as one of the most nutritious traditional leafy vegetables due to its high protein content and its richness in lysine, which is usually a limiting amino acid in cereals [6]. Amaranth has nutritional benefits, and it can be used in Nutritional Value Bakery Products. [7].

Amaranthus (family *Amaranthaceae*) collectively known as amaranth is a cosmopolitan genus of annual or short-lived perennial plants consisting of approximately 60 species. These species can be divided into cereals and vegetable amaranths based on their use for human consumption [8]. *Amaranthus* sp. is one of the neglected leafy vegetables. Amaranth is rich in protein, fiber, and minerals, with higher contents than cereals [9]. However, amaranth is affected by many factors that limit its production. Some of the major constraints on the production of leafy vegetables include poor quality seed, lack of production technologies, and poor marketing and processing strategies [10]. Low research investment is due to the status of the indigenous plant. Pests and diseases of Amaranth also pose serious threats to its sustainable production. Leaf miners, leaf rollers, cutworms, aphids, and flea beetles [11]. Severe infestations can lead to complete yield loss as the leaves become unsuitable for marketing and consumption [12]. Several research studies have been published on various aspects of amaranth with the aim of valorizing it and showing its importance, including its transformation and use in Guatemala, Mexico, and Peru [13], the development of genetic material and agronomic surveys in Mexico [14], and the nutritional and anti-nutritional composition of amaranth leaf vegetables in India [15]. A comprehensive analysis of published results on *Amaranthus* spp., including aspects such as composition, and antioxidant properties. Applications and processing were conducted by [16]. The work of [17] in Belgium provided insights into seed characteristics chemical composition, and technological properties of amaranth. Despite these studies, amaranth remains relatively unknown and neglected in the Sahelian region of Africa. This limited awareness likely contributes to the low collection and understanding of its genetic

variation in this area. African leafy vegetables like amaranth, can be grown under harsh climatic conditions with low external inputs. It is therefore important to explore the possibility of increasing production and marketing such vegetables in Mali. This will not only contribute to improved livelihoods of poor communities, but it will also indirectly contribute to improved nutrition and conservation of these valuable genetic resources. In efforts to be sustainable, it is important to understand the diversity of Amaranth in Mali in order to produce, market, and conserve it. For an indigenous crop with little information, a comparison of various accessions would be an important step in the identification of those morphological characteristics. According to various reports, the genus of amaranth exhibits a high degree of morphological diversity [11] [18] [19]. The interplay between local environmental conditions and the agro-morphological characteristics of Amaranth in Mali underscores the importance of context-specific agricultural practices. Understanding these relationships can aid in selecting suitable accessions for cultivation that are resilient to local climatic challenges while maximizing nutritional benefits and food security. This knowledge is vital for promoting sustainable agricultural practices that leverage local biodiversity and traditional knowledge systems.

Research indicates that local climatic conditions, such as temperature and rainfall patterns, significantly affect the growth and yield of Amaranth, influencing traits like plant height and leaf size [20]. Furthermore, soil characteristics, including nutrient availability and pH levels, play a crucial role in determining the morphological traits of Amaranth plants, as fertile soils generally support better growth outcomes [21]. Additionally, water availability is critical; regions with consistent rainfall or effective irrigation systems tend to produce healthier plants with superior morphological characteristics [22]. The importance of local agricultural practices cannot be overstated, as traditional methods such as crop rotation can enhance soil health and biodiversity, positively impacting plant growth [23]. Lastly, the genetic diversity of Amaranth accessions allows for better adaptation to specific local conditions, contributing to resilience against environmental stresses [20]. Such information is important for selecting better accession for breeding programs. Breeders and farmers need agro-morphological characterization and a lot of information and amaranth accessions efficient for increasing productivity [18]. So far, no distinctive study has been undertaken in Mali to characterize amaranth accessions. The current study was undertaken to characterize amaranth accessions using agro-morphological traits under agro-climatic conditions of Samanko. The results are useful in future genetic studies to improve this climate-resilient and highly nutritive underutilized crop species.

2. Materials and Methods

2.1. Experimental Site

The assessment was carried out during the 2023 rainy season in Mali, specifically at the World Vegetable Center experimental station in Samanko (**Figure 1**). This

site receives an annual rainfall of 1047 mm and features nutrient-rich clay soil with high water retention capacity but it can be difficult to work with, especially when dry. The average temperature at the site is 27.7°C. **Figure 1** illustrates the location of the experimental site and the general distribution of rainfall in this area

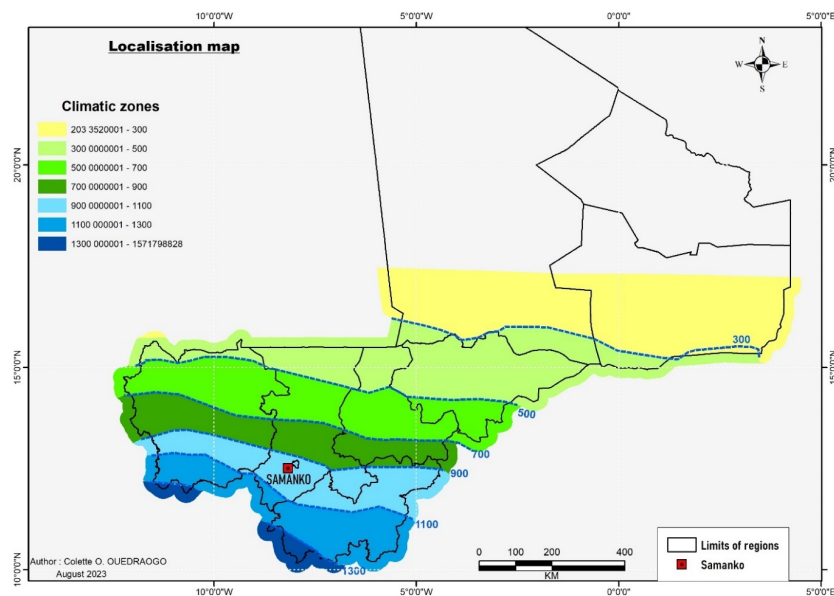


Figure 1. Map of the study area.

2.2. Amaranth Accessions Collections

Ten amaranth accessions were collected from different zones in Mali.

2.3. Experimental Design

A set of ten amaranth accessions were evaluated in a single environment using a randomized complete block design with three replications. The randomization of the experimental design was facilitated using the Breeding Management System (BMS) software. Each experimental plot consisted of lines measuring 5 meters in length. Planting densities were set at 50 cm × 50 cm between and within rows, respectively.

2.4. Data Collection

Data on both quantitative and qualitative traits were collected. The quantitative traits included plant height (cm) (from ground level to the highest tip of the inflorescence) measured using a measuring tape, stem girth (cm) (between the 3rd and 4th node) measured using a caliper, leaf length (cm), leaf width (cm), and inflorescence length (cm) measured using a measuring tape. The days to 50% flowering were determined by counting the days from planting to when 50% of the plants in the net plot flowered. Qualitative traits such as inflorescence color, plant growth habit, leaf pigmentation, petiole pigmentation, flower density index, stem color, inflorescence spininess, seed color, and the attitude of the terminal inflorescence were

visually observed on all plants within the gross plot.

2.5. Data Analysis

XLSTAT 2014.5.03 software was used for the analysis of variance. The Fisher test was used for the comparison of means when the analysis of variance revealed significant differences among the genotypes at a 5% probability threshold [24] [25].

Principal component analysis (PCA) was performed to reduce the dimension of the variables, to study the variability within the plant material, and to determine the different groups derived from the analysis. The multidimensional statistical method used to analyze the relationships between agromorphological parameters and yield components was principal component factor analysis (PCA), conducted using XLSTAT 2014.5.03 software on centered-reduced data. Thirty-two agromorphological parameters and yield components were analyzed, with these parameters serving as active or explanatory variables in the Principal Component Analysis (PCA). The variables included the number of plants raised, growth habit, main stem height, branching index, the average length of basal branches, average length of terminal branches, stem pigmentation, leaf length, basal branch length, length of the terminal branches; leaf length; leaf width; flowering date; length of the terminal inflorescence of the stem, length of the terminal inflorescence of the branch, weight of 1000 grains and length of axillary inflorescence, leaf pigmentation, prominence of the veins, attitude of the terminal inflorescence.

Additionally, ten varieties were used as supplementary variables: “Madira 1” “Madira 2.” “A2004” “A2002”, “N’gourouma”, “Akeri”, “AC-NL”, “Poly” and “AHTI”. An ascending hierarchical classification (CAH) was applied to construct groups of varieties according to agro-morphological characteristics and yield components using XLSTAT 2014.5.03 software.

3. Results

3.1. Agro-Morphological Characteristics of Amaranth Accessions

3.1.1. Qualitative Characteristics

Variability for morphological traits and their frequency distribution

Variability for morphological characters and their frequency distribution is shown in **Table 1**.

Growth habit: it was observed that 60% of the accessions studied produced an erect growth habit with 40% showing a semi-erect growth habit.

Stem and leaf pigmentation: Stem and leaf colour of the study accessions (60%) were mainly green.

Petiole pigmentation and Inflorescence Pigmentation: Accessions with white flowers had mostly green stems (60%) and low petiole pigmentation (53.3%).

Flower Density Index: half of the accessions had an average flower density index (50%).

Branching index: more than half of the accessions (60%) had erected ports.

Seed color: the lovers presented three colors of seeds.

Stem pubescence: half of the accessions had pubescent stems.

Branching index (IR): Branching index score ranged from 1 to 3. The IR score ranged from 1 to 3. Sixty percent of the accessions had an IR score of 3.

Table 1. Variability for morphological traits and their frequency distribution in amaranth in Samanko conditions.

Characteristic	Particulars/states	Descriptor score	Number of accessions	Proportion (%)
Growth habit	Faible	1	6	60
	Forte	2	4	40
	Absent	0	5	50
Stem pubescence		3	4	40
		7	1	10
Stem pigmentation	Absent	1	6	60
	Present	2	4	40
		6	1	10
Leaf pigmentation		7	1	10
		8	6	60
		9	2	20
Petiole pigmentation		1	5	53.3
		2	4	36.7
		3	1	10
Inflorescence Pigmentation		1	6	60
		2	3	30
		4	1	10
Flower Density Index		2	1	10
		3	1	10
		5	5	50
Branching index		7	3	30
		1	1	10
		2	3	30
Seed color		3	6	60
		1	5	50
		4	2	20
		5	3	30

Principal coordinate analysis

The four main axes (axis 1, 2, 3 and 4) explained 35.073; 20.94; 12.402 and 11.415% of the total variation respectively for the accessions giving a cumulative total variation of 79.831% (Table 1). Axes 1, 2, 3 and 4 contributed 5.962, 3.660, 2.108 and 1.940 of the eigenvalues respectively, grown accessions at Samanko conditions (Table 2).

Thorn in the leaf axil, Seed Shape, Seed Shell Type, Seed color, Pod dehiscence, and Flower Type were the main traits that contributed positively to PC1 for the accessions (Table 2). It was also observed that Axis 1 is strongly associated with traits such as the attitude of the terminal inflorescence ($r = 0.003$), the shape of the terminal inflorescence ($r = 0.0001$), and the shape of the leaves ($r = 0.005$).

Attitude of the terminal inflorescence; Presence of axillary inflorescence, Shape

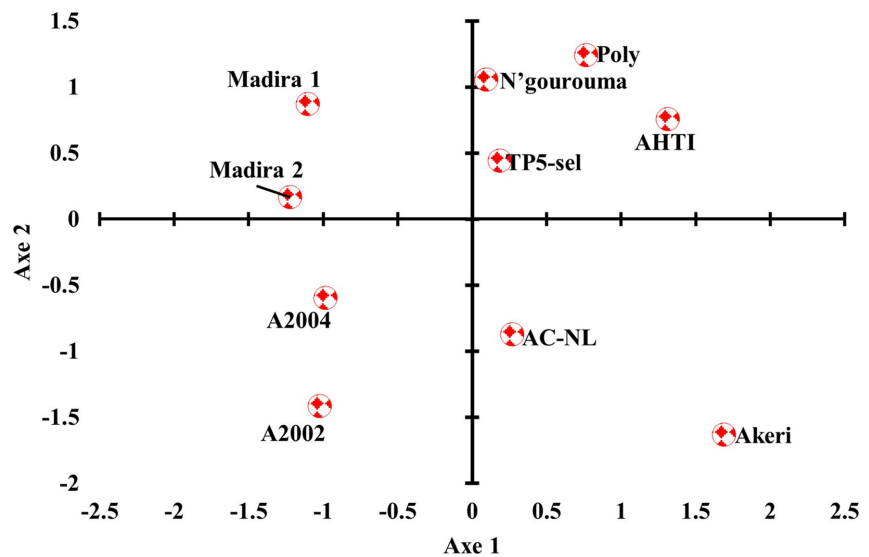
of the edge of the limb, Growth habit, Flower two characters Presence of axillary inflorescence Shape of the edge of the limb were important for PC3. Petiole pigmentation, stem pigmentation, pubescence of the stem and branching index contributed positively to PC4. Density Index was the most important character that contributed to the second principal component. Nearly all the characters that made significant contributions to a particular principal component were important contributors to another principal component (**Table 2**).

Table 2. Eigenvalues^a, eigenvectors^b and percentage of variation explained by the first four principal components for 10 amaranth accessions.

Qualitative character	Principal components			
	F1	F2	F3	F4
Petiole pigmentation	0.192	0.054	0.055	0.400
Attitude of the terminal inflorescence	0.003	0.802	0.039	0.014
Terminal Inflorescence shap	0.000	0.011	0.728	0.165
Presence of axillary inflorescence	0.262	0.331	0.019	0.079
prominent ribs	0.006	0.075	0.807	0.016
Shape of the edge of the limb	0.000	0.726	0.001	0.025
Leaf shap	0.005	0.022	0.013	0.939
Leaf pubescence	0.031	0.030	0.063	0.112
Leaf pigmentation	0.281	0.387	0.201	0.000
Thorn in the leaf axil	0.000	0.000	0.000	0.000
Stem pigmentation	0.023	0.005	0.036	0.904
Pubescence of the stem	0.170	0.010	0.140	0.533
Growth habit	0.042	0.422	0.005	0.282
Branching index	0.217	0.169	0.034	0.349
Seed Shape	0.000	0.000	0.000	0.000
Seed Shell Type	0.840	0.004	0.005	0.083
Seed color	0.820	0.004	0.002	0.052
Pod dehiscence	0.900	0.022	0.006	0.002
Flower Density Index	0.184	0.371	0.026	0.015
Flower Type	0.000	0.000	0.000	0.000
<i>Eigenvalue</i>	5.962	3.560	2.108	1.940
<i>Variabilition explained (%)</i>	35.073	20.941	12.402	11.415
<i>Cumulative %</i>	35.073	56.014	68.416	79.831

^aEigenvalues indicate the amount of variance explained by each principal component; ^bEigenvectors are the weights in a linear transformation when computing principal components; ^cValues in bold correspond for each variable to the factor for which the squared cosine is the largest.

Figure 2 shows the genetic relationships based on the first (axis 1 and 2) and second (axis 1 and 3) planes of the PCoA. Generally, the separation between accessions was high, with the first and second planes clearly separating accessions. The accessions were also clearly separated by the second plane (axis 1 and 3) and the first plane (axis 1 and 2) (**Figure 2**).



Red font point: accessions

Figure 2. Biplot analysis of axis 1 and 2 of principal coordinate analysis of 10 amaranth accessions based on dissimilarity of the qualitative characters.

3.1.2. Quantitative Characteristics

Table 3(A) and **Table 3(B)** reports agro-morphological parameters of Amaranth accessions such as: Number of plants (NP), stem height (HT), Basal branch length (LMBLB), length of the terminal branches (LMBLT), Leaf length (LoF), Leaf width (LaF), Flowering date (DF50%), Length of the terminal inflorescence of the stem (LITT), Length of the terminal inflorescence of the branch (LITB), Axillary inflorescence length (LIA) and Weight of 1000 grains (P1000).

Number of plants (NP) and stem height (HT): The statistical analysis showed a significant difference between accessions for the number of plants and stem height at the 5% level. The AHTI accession recorded the largest number of plants, followed by A2004. Madira 1. Madira 2, and N'gourouma. The accession Poly recorded the lowest number of plants. The average number of plants was 60 ± 5.8 , ranging from 34 to 64. Stem height varied from 95.5 cm to 247.2 cm. with an average of 159.04 ± 33.45 cm. The Madira 2 and Madira 1 accessions exhibited significantly greater plant heights, with Akeri accessions following closely behind, showing no statistically significant difference from these two accessions.

Basal branch length (LMBLB) and length of the terminal branches (LMBLT): The mean basal branch length (LMBLB) ranged from 26.6 cm to 126.6 cm. with an average of 59.43 ± 22.69 cm. while the average length of the terminal branches (LMBLT) varied between 8.7 cm and 41 cm. with an average of 23.05 ± 7.87 cm. Statistical analysis showed a significant difference in both LMBLB and LMBLT among the accessions at the 5% level. However, no significant difference was observed between the Akeri, AHTI, Poly, and TP5-sel accessions, all of which recorded the highest values for LMBLB and LMBLT. On the other hand, Nadira 1 and Madira 2 accessions recorded the lowest LMBLB and LMBLT, with no significant difference compared to the A2004, A2002, and AC-NL accessions for these

variables.

Leaf length (LoF) and Leaf width (LaF): The statistical analysis showed significant differences between amaranth accessions for leaf length (LoF) at the 5% level. The LoF varied from 4.75 cm to 16.3 cm. with an average of 12.3 ± 2.76 cm. The Madira 1 accession recorded the highest value. followed by Madira 2, N'gourouma, and A2004, which were statistically similar. Highly significant differences were observed among the accessions for leaf width (LaF) at the 5% level. The LaF ranged from 3.9 to 10.7 cm. with an average of 6.9 ± 1.84 cm. The TP5-sel and N'gourouma varieties recorded the highest LaF values, followed by the Poly variety. In contrast. Madira 1 and A2002 had the lowest LaF measurements, with no statistically significant difference between them.

Flowering date (DF50%): The 50% flowering date ranged from 40 to 60 days. with an average of 50 ± 6.26 days. Akeri, Madira 2, A2004. Madira 1, AC-NL and A2002 all exhibited statistically similar results, recording the longest time to reach 50% flowering. These were followed by the N'gourouma variety. AHTI. Poly. and TP5-sel accessions recorded the shortest time to reach 50% flowering compared to the other accessions.

Length of the terminal inflorescence of the stem (LITT) and Length of the terminal inflorescence of the branch (LITB): The average length of the terminal inflorescence of the stem (LITT) was 5.609 ± 5.28 cm. with a range of 1.3 to 21.3 cm. The length of the terminal inflorescence of the branch (LITB) varied from 3.2 to 22 cm with an average of 8 ± 4.849 cm. Statistical analysis showed a highly significant difference between accessions for both LITT and LITB. AHTI and the accession Poly recorded the highest LITT, followed by Akeri and TP5-sel accessions, which were statistically similar. The lowest LITT values were recorded by Madira 1. Madira 2, N'gourouma. A2004, A2002 and AC-NL which were statistically equal. The Poly variety recorded the highest LITB, followed by AHTI, Akeri and TP5-sel accessions. No significant difference was noted between A2004, Madira 1, AC-NL, A2002, N'gourouma and Madira 2 accessions which recorded the lowest LITB values.

Axillary inflorescence length (LIA): Statistical analysis showed a highly significant difference in LIA among the accessions at the 5% significance level. Axillary inflorescence length varied from 2.35 cm to 17.8 cm. with an average of 7.23 ± 4.65 cm. AHTI and Poly accessions recorded the highest LIA averages of 14.76 and 14.56 cm. respectively. They were followed by Akeri and TP5-sel accessions which were statistically similar. The lowest values were recorded by N'gourouma, Madira 1, Madira 2, A2002, A2004 and AC-NL which were statistically similar.

Weight of 1000 grains (P1000): The weight of 1.000 grains ranged from 0.83 to 1.57 g (± 0.213 g), with an average weight of 1.11 g. Statistical analysis at the 5% significance level indicated a highly significant difference between the varieties. The AHTI and Poly varieties had the highest average 1.000-grain weights and were statistically similar. The Akeri variety followed them. In contrast, the N'gourouma. A2002, A2004, Madira 1 and AC-NL varieties recorded the lowest 1000grain weights.

Table 3. (A). Agro-morphological parameters of Amaranth accessions. (B). Agro-morphological parameters of Amaranth accessions.

(A)						
Amaranth accessions	NPL	Htige	LMBLB	LMBLT	LoF	LaF
Akeri	56.66 ± 4.04 ^{ab}	180.56 ± 13.4 ^{ab}	57.767 ± 22.31 ^{abc}	30.767 ± 4.17 ^a	13.356 ± 1.22 ^{abc}	7.47 ± 0.87 ^c
AHTI	64.00 ± 0.01 ^a	136.53 ± 3.11 ^c	62.70 ± 6.39 ^{abc}	30.233 ± 9.66 ^a	11.917 ± 1.15 ^{bc}	6.95 ± 1.1 ^c
Poly	49.33 ± 13.42 ^b	125.43 ± 26.54 ^c	87.33 ± 34.35 ^a	32 ± 5.02 ^a	8.8 ± 4.29 ^d	9.26 ± 0.5 ^{ab}
N'gourouma	62.67 ± 0.57 ^a	179.05 ± 10.88 ^{ab}	84.43 ± 14.19 ^{ab}	24.83 ± 1.24 ^{ab}	12.917 ± 1.17 ^{bc}	9.53 ± 1.12 ^a
Madira 2	62 ± 2.64 ^a	210.2 ± 52.59 ^a	53.167 ± 20.46 ^{bc}	14.07 ± 5.91 ^c	14.41 ± 0.86 ^{ab}	6.55 ± 0.17 ^c
TP5-sel	59.33 ± 4.04 ^a	143.13 ± 9.82 ^{bc}	67.50 ± 18.2 ^{abc}	28.47 ± 4.52 ^a	10.28 ± 4.84 ^{cd}	10 ± 2.13 ^a
A2004	63.33 ± 1.15 ^a	147.43 ± 24.04 ^{bc}	43.53 ± 13.65 ^{cd}	18.48 ± 7.19 ^{bc}	12.08 ± 0.9 ^{abcd}	6.01 ± 1.14 ^{cd}
Madira 1	61 ± 3 ^a	188.13 ± 3.65 ^a	36.96 ± 16.17 ^c	16.40 ± 0.95 ^{bc}	15.97 ± 0.29 ^a	4.28 ± 0.33 ^d
AC-NL	61 ± 2.64 ^a	146.46 ± 25.52 ^{bc}	40.4 ± 9.70 ^c	18.03 ± 3 ^{bc}	12.56 ± ^{abcd}	6.56 ± 0.85 ^{bc}
A2002	58.67 ± 0.57 ^a	133.46 ± 19.07 ^c	60.56 ± 17.09 ^{abc}	17.26 ± 0.96 ^{bc}	10.58 ± 1.94 ^{bcd}	4.76 ± 0.69 ^d
<i>Pr > F</i>	0.055	0.003	0.041	0.001	0.043	< 0.0001
<i>Signification</i>	<i>S</i>	<i>S</i>	<i>S</i>	<i>S</i>	<i>S</i>	<i>THS</i>
<i>CV (%)</i>	7	18	29	29	24	17
(B)						
Amaranth accessions	DF50%	LITT	LITB	LIA	P1000	
Akeri	54 ± 0.01 ^a	6.80 ± 1.47 ^b	11.40 ± 0.52 ^b	7.917 ± 1.45 ^{bc}	1.34 ± 0.03 ^b	
AHTI	40 ± 0.01 ^d	13.27 ± 2.75 ^a	13.13 ± 2.26 ^b	14.76 ± 0.66 ^a	1.4 ± 0.07 ^{ab}	
Poly	42.33 ± 4.04 ^{cd}	15.96 ± 5.2 ^a	17.03 ± 4.3 ^a	14.56 ± 3.59 ^a	1.47 ± 0.11 ^a	
N'gourouma	49.33 ± 4.04 ^b	2.967 ± 0.47 ^c	4.98 ± 0.75 ^c	5.81 ± 2.24 ^{cd}	0.93 ± 0.04 ^{ef}	
Madira 2	57.33 ± 2.3 ^a	2.643 ± 1.45 ^c	4.25 ± 0.54 ^c	3.46 ± 0.99 ^c	0.97 ± 0.04 ^{def}	
TP5-sel	45 ± 1.15 ^c	6.78 ± 2.51 ^b	11.23 ± 2.83 ^b	10.07 ± 2.99 ^b	1.67 ± 0.0 ^{4c}	
A2004	54.66 ± 1.15 ^a	1.85 ± 0.2 ^c	4.717 ± 0.95 ^c	4.45 ± 1.62 ^d	0.947 ± 0.12 ^{ef}	
Madira 1	55.33 ± 1.15 ^a	1.967 ± 0.47 ^c	4.80 ± 1.05 ^c	5.23 ± 2.08 ^{cd}	0.90 ± 0.01 ^e	
AC-NL	55.33 ± 1.15 ^a	1.78 ± 0.56 ^c	3.62 ± 0.36 ^c	2.65 ± 0.36 ^d	1.07 ± 0.04 ^{def}	
A2002	54 ± 0.01 ^a	2.05 ± 0.68 ^c	4.82 ± 0.81 ^c	3.36 ± 1.26 ^d	1.03 ± 0.05 ^d	
<i>Pr > F</i>	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	
<i>Significant</i>	<i>THS</i>	<i>THS</i>	<i>THS</i>	<i>THS</i>	<i>THS</i>	
<i>CV (%)</i>	12	92	60	62	19	

Means followed by the same letters in a column are not statistically different at $P < 0.05$. Abbreviation: LITT: Length of the inflorescence terminal of the stem (cm); LITB: Length of the terminal inflorescence of the branch (cm); DF 5%: Date 5% flowering; P1000: Weight of 1000 grains (g); LIA: Length of inflorescence Axillary (cm).

3.1.3. Correlation Analysis between Variables Quantitative and Accessions

The correlation between the different variables is presented in **Table 4**. There were significant ($P < 0.05$) and positive correlations between most of the quantitative variables (**Table 4**). The correlation analysis reveals highly significant correlations. From the conformity test of the correlation coefficients. It appears that the Axillary Inflorescence (LIA) and the Length of the Terminal Inflorescence are correlated ($r = 0.82$). Length of the inflorescence terminal of the stem (LITT) and the Length of the Terminal Basal Branches (LITBT) are correlated ($r = 0.75^*$). Leaf length (LoF) is correlated with stem height ($r = 0.36^*$). The 50% flowering date presents a very strong correlation with stem height ($r = 0.447^*$). Length of the

inflorescence terminal of the stem was highly correlated with Leaf length ($r = 0.414^*$). Length of inflorescence Axillary was also highly correlated with Length of the inflorescence terminal of the stem ($r = 0.904^*$). A negative and very highly significant correlation ($r = -0.824$) emerges between the Length of the Axillary Inflorescence (LIA) and the 50% flowering date. There is also a negative correlation between the weight of 1000 grains and the 50% flowering date ($r = -0.664^*$). Meanwhile in Length of the inflorescence terminal of the stem and stem height recorded a highly significant ($p < 0.05$) negative correlation ($r = -0.393^*$) as well as between Leaf length and Date 50% flowering ($r = -0.466^*$) and between sheet width and Length of the terminal inflorescence of the branch ($r = -0.459^*$).

Table 4. Relationship among variables.

Variables	NPL	Htige	LMBLB	LMBLT	LoF	LaF	DF50%	LITT	LITB	LIA	P1000
NPL	1										
Htige	0.092	1									
LMBLB	-0.181	-0.054	1								
LMBLT	-0.207	-0.234	0.493*	1							
LoF	0.369	0.639	-0.071	-0.336	1						
LaF	-0.269	-0.072	0.466*	0.526	-0.394	1					
DF50%	0.229	0.447	-0.424	-0.635	0.470	-0.466	1				
LITT	-0.223	-0.393	0.415	0.751	-0.420	0.414	-0.808	1			
LITB	-0.337	-0.400	0.375	0.728	-0.459	0.443	-0.790	0.922	1		
LIA	-0.138	-0.328	0.517	0.737	-0.222	0.423	-0.824	0.904	0.849	1	
P1000	-0.323	-0.396	0.349	0.739	-0.372	0.438	-0.664	0.874	0.902	0.839	1

*Correlation is significant at the $P > 0.05$ level. Abbreviation: NPL: Number of plants raised; LITT: Length of the inflorescence terminal of the stem; HT: stem height; LMBLB length of basal Branches; LMBLT: Length of terminal branches; LITB: Length of the terminal inflorescence of the branch; DF50%: Date 50% flowerin; LIA: Length of inflorescence Axillary; LaF: sheet width; LoF: Leaf length, P1000: Weight of 1000 grains.

3.1.4. Principal Component Analysis

The principal component analysis is presented in **Table 5**. To capture the majority of the information in **Table 5**. It is necessary to consider the third and fourth axes, which together account for 80.07% of the total variability

- **Axis 1**, which explains 43.04% of the variability, is primarily influenced by the variable's presence of axillary inflorescence, length of the terminal inflorescence of the branch, length of the terminal inflorescence of the stem, shape of the leaf, length of the terminal branches and Weight of 1000 grains. This axis contrasts varieties with a prominent Length of inflorescence Axillary, Terminal inflorescence of the branch, Length of the inflorescence terminal of the stem, 50% flowering date, Leaf pubescence, length of terminal branches, Number of plants raised, Growth habit, Stem height, Weight of 1000 grains, Seed Shape Seed Shell Type, Seed color and significant pod dehiscence.
- **Axis 2**, which accounts for 15.33% of the variability, is primarily explained by the variable related to Terminal Inflorescence shap, Shape of the edge of the limb, Branching index, Pod dehiscence and Flower Density Index.
- **Axis 3**, which accounts for 11.804% of the variability, is characterized by the

variables Terminal Inflorescence shap, presence of axillary inflorescence and Leaf length.

- **Axis 4**, which accounts for 9.903% of the variability, is characterized by the variables petiole pigmentation, length of basal branches, sheet width, stem pigmentation and pubescence of the stem.

Figure 2 shows correlations between the variables and the factors. The first axis accounts for 43.04% of the total variation, while the second axis explains 15.33% of the total variability.

Table 5. Correlations between variables and factors.

Characters	Principal component			
	F1	F2	F3	F4
Petiole pigmentation	0.017	0.158	0.011	0.656
Length of inflorescence Axillary	0.950	0.000	0.019	0.008
Attitude of the terminal inflorescence	0.176	0.405	0.314	0.002
Terminal Inflorescence shap	0.077	0.114	0.350	0.000
Terminal inflorescence of the branch	0.858	0.011	0.001	0.043
Presence of axillary inflorescence	0.095	0.000	0.379	0.205
Length of the inflorescence terminal of the stem	0.903	0.000	0.000	0.027
50% flowering date	0.903	0.024	0.001	0.005
Length of basal branches	0.290	0.187	0.078	0.308
prominent ribs	0.082	0.014	0.538	0.242
Shape of the edge of the limb	0.038	0.328	0.178	0.023
Shape of the leaf	0.321	0.000	0.027	0.487
Leaf pubescence	0.151	0.079	0.032	0.002
Leaf pigmentation	0.000	0.834	0.044	0.000
Sheet width	0.210	0.081	0.007	0.618
Leaf length	0.423	0.021	0.541	0.004
Thorn in the leaf axil	0.000	0.000	0.000	0.000
Stem pigmentation	0.335	0.010	0.041	0.447
Pubescence of the stem	0.157	0.053	0.067	0.485
length of terminal branches	0.693	0.043	0.001	0.176
Number of plants raised	0.206	0.004	0.165	0.086
Growth habit	0.600	0.166	0.006	0.075
Stem height	0.399	0.005	0.377	0.074
Branching index	0.001	0.473	0.050	0.407
Weight of 1000 grains	0.745	0.104	0.010	0.057
Seed Shape	0.000	0.000	0.000	0.000
Seed Shell Type	0.487	0.231	0.007	0.050

Continued

Seed color	0.487	0.258	0.000	0.015
Pod dehiscence	0.120	0.505	0.006	0.011
Flower Density Index	0.086	0.373	0.236	0.000
Flower Type	0.000	0.000	0.000	0.000
<i>Eigenvalue</i>	12.480	4.447	3.423	2.872
<i>Variability explained (%)</i>	43.036	15.334	11.804	9.903
<i>Cumulative %</i>	43.036	58.370	70.174	80.077

^aEigenvalues indicate the amount of variance explained by each principal component; ^bEigenvectors are the weights in a linear transformation when computing principal components; ^cValues in bold correspond for each variable to the factor for which the squared cosine is the largest.

3.2. Ascending Hierarchical Classification (AHC)

Figure 3 illustrates the dendrogram showing the relationships among the ten amaranth accessions. The hierarchical clustering analysis (HCA) revealed three distinct clusters: C1, C2 and C3.

- **Class 1 (C1)** is located in the negative part of Axis 1 and is characterized by several key features. Varieties in this class have a long flowering period, with 50% of plants reaching flowering at an average of 55 days. The average stem height is 169.58 cm. and leaf length measures 13.33 cm. These varieties also exhibit a good attitude of terminal inflorescence. The accessions classified under C1 include “Madira 1”. “Madira 2”, “A2004” and “A2002”. Additional Traits: These accessions have petioles with more or less pigmentation. Their leaves are more than 2 cm longer than those of the third class and they are 36.6 cm taller than Class 3 varieties.
- **Class 2 (C2)** is situated in the central part of the two axes and exhibits the following characteristics: The flowering date is relatively average with 50% of plants flowering in about 51 days. The average stem height is 162.304 cm. and leaf length measures 12.28 cm. Varieties in this class are noted for their prominent veins and strong stem pigmentation. Accessions in C2 include “N’gourouma”, “Akeri”, “TP5-sel” and “AC-NL”. Additionally these accessions have average 1000 grains weights and basal branch lengths of 13 cm. Their leaf widths are 2 cm greater than those in Class 1 and 1 cm less than those in Class 3.
- **Class 3 (C3)** is located in the positive part of Axis 1 and is characterized by the following traits: The accessions in this class include “Poly” and “AHTI”. Notable features are the lengths of inflorescences, such as axillary inflorescence and terminal inflorescence of the stem. The leaf shape is described as good and the growth habit is strong. Varieties in this class have a high 1000 grains weight and well-represented seed hulls. They exhibit average basal and terminal branch lengths of 31.11 cm and 75 cm. respectively. Additionally the flowering date is notably shorter at 41 days for 50% flowering and the average stem height is 130.98 cm. Additional Traits: These accessions have 1.5 times more weight of 1000 grains compared to Class 1 accessions.

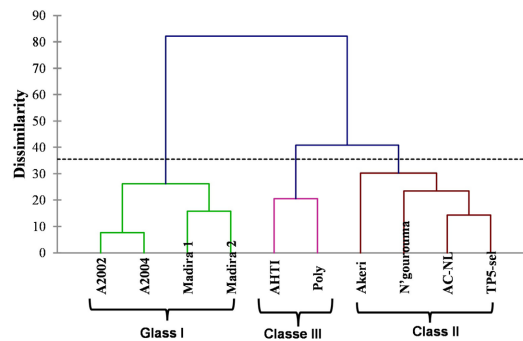


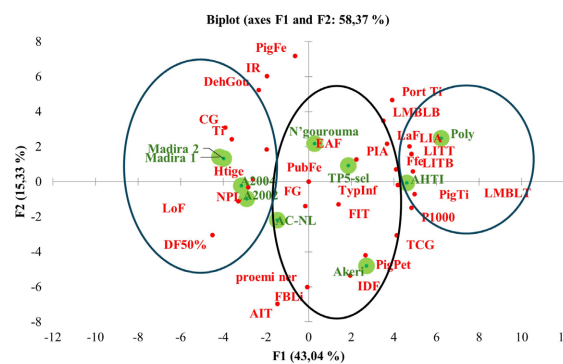
Figure 3. Dendrogram showing the relationship among the ten accessions.

3.3. Relationship between Variables and Accessions

Figure 4 highlights the relationships between the different yield component variables and the accessions. The circle of correlations between variables and the genotypes in the main plan 1 - 2.

• **Close Relationships:**

- Accessions “Akeri”, “TP5-se”, “AC-NL” and “N’gourouma” are closely associated with several variables: the weight of 1000 grains (P1000), stem pigmentation (PigTi). petiole pigmentation, average length of the terminal branches (LITBT), and the and the shape of the leaf (Ffe).
- Accessions “Poly” and “AHTI” are also closely related. They are associated with the average length of basal branches (LMBLB), growth habit (Por ti), length of the terminal inflorescence of the stem (LITT), and length of the terminal inflorescence of the branch (LITB), presence of axillary inflorescence (LIA).



Red font: character; Green font: accessions. Abbreviation: NPL: Number of plants raised; LITT: Length of the inflorescence terminal of the stem; HT: stem height; LMBLB length of basal branches; LMBLT: Length of terminal branches; LITB: Length of the terminal inflorescence of the branch; DF50%: Date 50% flowering; LIA: Length of inflorescence Axillary; LaF: sheet width; LoF: Leaf length. Ti: Petiole pigmentation, AIT: Attitude of the terminal inflorescence, FIT: Terminal Inflorescence shap, PIA: Presence of axillary inflorescence, proemi ner: prominent ribs, FBLi: Shape of the edge of the limb, Ffe: Leaf shap, FBLi: Leaf pubescence, PigFe: Leaf pigmentation, EAF: Thorn in the leaf axil, PigTi: Stem pigmentation, FBLi: Pubescence of the stem, Port Ti :Growth habit, IR: Branching index, FG: Seed Shape, TypInf: Seed Shell Type, CG: Seed color, DehGou: Pod dehiscence, IDF: Flower Density Index, TypInf: Flower Type. P1000: Weight of 1000 grains.

Figure 4. Relationship between amaranth accessions and variables.

Furthermore, the position of the accessions “Madira 1”; “Madira 2”, “A2004”, “A2002” on the correlation circle indicates that the height of the plants, the number of plants, the length of the leaves, the 50% flowering date, the color of the pods, the pigmentation of the stems are among the quantitative and qualitative characters those which explain the variability of the accessions the most.

4. Discussion

The major objective of this study was to assess the genetic diversity within the collection to understand the range of genetic variation [20]. The study of [5] revealed a significant level of morphological diversity between and within amaranth accessions from Malawi. Significant variation was observed for all the agro-morphological parameters among the ten amaranth accessions from Samako in Mali. This wide range of variation for qualitative and quantitative traits is consistent with the findings of [5] and [26]. Overall, there was a clear separation of the field grown accessions from Samanko conditions based on the dissimilarity of the qualitative characters. This diversity is due to amaranths variability in mating behavior and genetic material, adaptability to different growing conditions, and a wide range of variability in sexual systems, from monoecy to dioecy. This could be due to character expression being more vivid in the Samanko field. [27] and [26] reported differences between accessions with respect to characters like stem colour, leaf colour and seed colour. [28] reported wide genetic diversity for qualitative traits among 18 amaranth germplasm collections. [29]-[31] also reported polymorphism for these traits among different amaranthus accessions. In the present study, the two qualitative traits, namely, dense inflorescence index and Leaf pigmentation, showed high variability among the species. The diversity of Eighteen Amaranth germplasm collections in the Philippines and found a wide genetic variability among them for qualitative traits reported by [28].

Traits such as the number of plants, leaf width, stem height, length of the terminal inflorescence of the stem, and axillary inflorescence length. 50% flowering and weight of 1000 grains exhibited significant variation. This variation is likely due to genetic differences among the accessions evaluated since they were grown in the same environment. Similar results were reported by [5]. This observation contrasts with [32], who attributed vegetative trait diversity to environmental factors. Traditional African vegetables such as *Amarant* sp, and spider plant, are tolerant of abiotic stresses such as low soil moisture and heat conditions. In a study conducted in South Africa, amaranth was found to be more heat tolerant [33]. For [34], temperature also affects the production of amaranth biomass, under high temperatures, amaranth produces more biomass than in cool times, which indicates that the local environment can influence amaranth production. Considering the various environmental conditions, explains the existence of a long list of diversity of amaranth in the area. Significant variations in days to 50% flowering also point to genetic differences among the accessions. This was reported by [35], who demonstrated that early flowering in amaranth is governed by a single gene, with

the dominant allele determining earliness in flowering. [36] suggested that late flowering could be advantageous for vegetable amaranths, providing farmers with a longer harvesting period for the leaves.

This section will help to understand the relationships between different agro-morphological traits and how they influence each other which is crucial for breeding and improving amaranth species.

Understanding the associations between agro-morphological parameters is crucial for the selection process. The present study found narrow differences in correlation coefficients indicating a strong inherent association among traits. The 1000 Grain Weight (P1000) exhibited a highly significant and positive association with the length of the terminal inflorescence of the stem suggesting the potential for simultaneous improvement of these traits. The highly significant and positive correlation between the number of days to 50% flowering and the height of the plant indicates that plants that have a long cycle are those that have a larger size. Indeed these plants, given the length of their cycle have more time for vegetative development unlike those whose cycle is short. A negative and highly significant correlation ($r = -0.82$) was observed between the length of the axillary inflorescence and the 50% flowering date. This indicates that plants that have a short cycle are those that have a greater Length of the Axillary inflorescence. Indeed, these plants given the length of their Axillary inflorescence have the advantage of having a large number of flowers and a good reproductive capacity. Additionally a negative correlation was found between the weight of 1000 grains and the 50% flowering date. This indicates that plants with a long cycle have light weights of 1000 grains. In a context of low rainfall, water requirements could not be met, which has consequences on grain filling. Hence light grain weights. Similar findings were reported by [5] and [37], who noted negative and significant associations between grain yield and traits such as leaf length, plant height, stem girth, days to 50% flowering, and leaf yield. This section will help to understand the relationships between different agro-morphological traits and how they influence each other which is crucial for breeding and improving amaranth species. Understanding the associations between agro-morphological parameters is crucial for the selection process.

The dendrogram indicated genetic diversity within and between cluster groups. It showed variations in quantitative characters such as stem height, leaf length, axillary inflorescence length, terminal inflorescence length, growth habit, high 1000 grain weights, and seed hulls, High variability in qualitative characters, such as recovery terminal inflorescence attitude, prominent veins, and strong stem pigmentation was also evident. The grouping of varieties into three clusters based on agro-morphological characters reflects the diversity of the sample studied. This study suggests that the genetic material has significant potential for improvement. Accessions with desirable traits can be crossed with other promising accessions to produce the desired hybrids. For [38], this difference between group amaranth may be due to their collection from diverse geographical zones.

Plant height, days to flowering, leaf length, and individual leaf width are all key contributors to amaranth's leaf yield, as are a variety of other yield elements. [39] supported similar findings. The observed high plant height in the 80% (C1 and C2) leaf yielder accession might be due to the inherent genetic variation, strong light competition, and partition of more assimilate for stem elongation. Similar findings were observed by [40] in amaranths and taller plants outcompete weeds more successfully than shorter ones [41]. Similarly, the leaf area is crucial in determining the yield [42].

The sixty percent (60%) of accessions (C2 and C3) with high 1000 grain weight and very good inflorescence had the greatest leaf width and leaf shape. These accessions probably had the best leaf area. In addition, these accessions had very high petiole and stem pigmentation. When water availability increased, plants were able to perform photosynthesis more efficiently [43]. The leaf area intercepts sunlight, absorbs CO₂ and inorganic nitrogen, and performs photosynthesis and biomass accumulation, among other factors, determining yield, as reported by [44].

5. Conclusion

This study highlighted the extent of quantitative and qualitative variations among the ten amaranth accessions. The observed differences in agro-morphological parameter correlation coefficients and the identification of three distinct classes (C1: Madira 1, Madira 2, A2004, A2002; C2: TP5-sel. N'gourouma. Akeri. AC-NL; C3: AHTI, Akeri) provide ample opportunities for plant breeders to improve amaranth production. The identified genetic diversity within these accessions offers valuable insights for future breeding programs aimed at enhancing the yield and quality of amaranth.

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Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] Hughes, J. (2009) Just Famine Foods? What Contributions Can Underutilized Plants Make to Food Security? *Acta Horticulturae*, **806**, 39-48.
<https://doi.org/10.17660/actahortic.2009.806.2>

- [2] Keatinge, J.D.H., Yang, R.-Y., d'A. Hughes, J., Easdown, W.J. and Holmer, R. (2011) The Importance of Vegetables in Ensuring Both Food and Nutritional Security in Attainment of the Millennium Development Goals. *Food Security*, **3**, 491-501. <https://doi.org/10.1007/s12571-011-0150-3>
- [3] Mungofa, N., Sibanyoni, J.J., Mashau, M.E. and Beswa, D. (2022) Prospective Role of Indigenous Leafy Vegetables as Functional Food Ingredients. *Molecules*, **27**, Article No. 7995. <https://doi.org/10.3390/molecules27227995>
- [4] Shayanowako, A., Morrissey, O., Tanzi, A., Muchuweti, M., Mendiondo, G., Mayes, S., *et al.* (2021) African Leafy Vegetables for Improved Human Nutrition and Food System Resilience in Southern Africa: A Scoping Review. *Sustainability*, **13**, Article No. 2896. <https://doi.org/10.3390/su13052896>
- [5] Nyasulu, M., Sefasi, A., Chimzinga, S. and Maliro, M. (2021) Agromorphological Characterisation of Amaranth Accessions from Malawi. *American Journal of Plant Sciences*, **12**, 1528-1542. <https://doi.org/10.4236/ajps.2021.1210108>
- [6] Grobelnik, M.S., Turinek, M., Jakop, M., Bavec, M., and Bavec, F. (2009) Nutrition Value and Use of Grain Amaranth: Potential Future Application in Bread Making. *Agriculture*, **6**, 43-53.
- [7] Gebreil, S.Y., Ali, M.I.K. and Mousa, E.A.M. (2020) Utilization of Amaranth Flour in Preparation of High Nutritional Value Bakery Products. *Food and Nutrition Sciences*, **11**, 336-354. <https://doi.org/10.4236/fns.2020.115025>
- [8] Grobelnik Mlakar, S., Turinek, M., Jakop, M., Bavec, M. and Bavec, F. (2010) Grain Amaranth as an Alternative and Perspective Crop in Temperate Climate. *Journal for Geography*, **5**, 135-145. <https://doi.org/10.18690/rg.5.1.4011>
- [9] Uriyapongson, J. and Rayas-Duarte, P. (1994) Comparison of Yield and Properties of Amaranth Starches Using Wet and Dry-Wet Milling Processes. *Cereal Chemistry*, **71**, 571-577.
- [10] Dinssa, F.F., Stoilova, T., Rouamba, A., Tenkouano, A., Ebert, W.A., Hanson, P., *et al.* (2013) Prospects and Challenges for Preserving and Mainstreaming Underutilized Traditional African Vegetables. *3rd International Conference on Neglected and Underutilized Species (NUS): For a Food-Secure Africa*, Accra, 25-27 September 2013, 275-282.
- [11] Mng'omba, S.A., Kwapata, M.B. and Bokosi, J.M. (2003) Performance of Grain Amaranth Varieties under Drought Stressed Conditions in Two Contrasting Agro-Ecological Areas in Malawi. *Acta Horticulturae*, **618**, 313-319. <https://doi.org/10.17660/actahortic.2003.618.36>
- [12] Stoilova, T., Dinssa, F.F., Ebert, A.W. and Tenkouano, A. (2015) The Diversity of African Leafy Vegetables: Agromorphological Characterization of Subsets of AVRDC's Germplasm Collection. *Acta Horticulturae*, **1102**, 67-74. <https://doi.org/10.17660/actahortic.2015.1102.7>
- [13] Chelang'a, P.K., Obare, G.A. and Kimenju, S.C. (2013) Analysis of Urban Consumers' Willingness to Pay a Premium for African Leafy Vegetables (ALVs) in Kenya: A Case of Eldoret Town. *Food Security*, **5**, 591-595. <https://doi.org/10.1007/s12571-013-0273-9>
- [14] Espitia, E. (1992) Amaranth Germplasm Development and Agronomic Studies in Mexico. *Food Reviews International*, **8**, 71-86. <https://doi.org/10.1080/87559129209540930>
- [15] Prakash, D. and Pal, M. (1991) Nutritional and Antinutritional Composition of Vegetable and Grain Amaranth Leaves. *Journal of the Science of Food and Agriculture*, **57**, 573-583. <https://doi.org/10.1002/jsfa.2740570410>

- [16] Venskutonis, P.R. and Kraujalis, P. (2013) Nutritional Components of Amaranth Seeds and Vegetables: A Review on Composition, Properties, and Uses. *Comprehensive Reviews in Food Science and Food Safety*, **12**, 381-412. <https://doi.org/10.1111/1541-4337.12021>
- [17] De Bock, P., Daelemans, L., Selis, L., Raes, K., Vermeir, P., Eeckhout, M., *et al.* (2021) Comparison of the Chemical and Technological Characteristics of Wholemeal Flours Obtained from Amaranth (*Amaranthus* sp.), Quinoa (*Chenopodium quinoa*) and Buckwheat (*Fagopyrum* sp.) Seeds. *Foods*, **10**, Article No. 651. <https://doi.org/10.3390/foods10030651>
- [18] Srivastava, R. (2015) Assessment of Morphological Diversity of Selected Amaranthus Species. *Journal of Global Science*, **4**, 2-4.
- [19] Modi, A. (2009) Growth Temperature and Plant Age Influence on Nutritional Quality of *Amaranthus* Leaves and Seed Germination Capacity. *Water SA*, **33**, 369-376. <http://www.wrc.org.za>
<https://doi.org/10.4314/wsa.v33i3.49118>
- [20] Ezin, V., Kpanougo, C.M.I. and Ahanchede, A. (2022) Genetic Diversity and Environmental Influence on Morphological and Yield Parameters of Maize in Benin. *Heliyon*, **8**, e09670. <https://doi.org/10.1016/j.heliyon.2022.e09670>
- [21] Mbosso, C., Boulay, B., Padulosi, S., Meldrum, G., Mohamadou, Y., Berthe Niang, A., *et al.* (2020) Fonio and Bambara Groundnut Value Chains in Mali: Issues, Needs, and Opportunities for Their Sustainable Promotion. *Sustainability*, **12**, Article No. 4766. <https://doi.org/10.3390/su12114766>
- [22] USAID (2016) Africa Research in Sustainable Intensification for the Next Generation (Africa Rising) Mid-Term Evaluation.
- [23] Malhotra, N., Sharma, P., Sood, H., Chandora, R., Arya, M., Rana, J.C., *et al.* (2022) Agro-morphological Characterization and Nutritional Profiling of Traditional Himalayan Crop Landraces for Their Promotion toward Mainstream Agriculture. *Frontiers in Plant Science*, **13**, Article 898220. <https://doi.org/10.3389/fpls.2022.898220>
- [24] Gomez, A.K. and Gomez, A.A (1984) Statistical Procedures for Agricultural Research. 2nd Edition, John Wiley & Sons. Incorporation.
- [25] Steel, R.G.D., Torrie J.H. and Dickey, D.A. (1997) Principles and Procedures of Statistics: A Biometrical Approach. McGraw Hill Book Co., 400-428.
- [26] Shah, L.R., Afroza, I.B., Khan, S.H. and Habib, M. (2018) Morphological Characterization of *Amaranthus* spp. under Temperate Environment Using NBPGR Descriptor. *Pharmacognosy and Phytochemistry*, **7**, 2716-2718.
- [27] Akaneme, F.I. and Ani, G.O. (2013) Morphological Assessment of Genetic Variability among *Amaranthus hybridus*. *World Applied Sciences Journal*, **28**, 568-577.
- [28] Gueco, L.S., Borrromeo, T. and De Guzman, C. (2016) Diversity in the Morphology of Amaranth (*Amaranthus* sp.) Germplasm Collection in the Philippines. *Asian Journal of Agriculture and Food Sciences*, **4**, 73-79.
- [29] Bayer Crop Science (2013) *Amaranthus hybridus*. <https://www.cropscience.bayer.com/en/Crop-Compendium/Pests-Diseases-Weeds/Weeds/Amaranthushybridus.aspx>
- [30] Gerrano, A.S., Van Rensburg, W.J., Mavengahama, S., Bairu, M., Venter, S. and Adebola, P.O. (2017) Qualitative Morphological Diversity of Amaranthus Species. *Journal of Tropical Agriculture*, **55**, 12-20.
- [31] Kamble, S.R. and Gaikwad, D.K. (2017) Qualitative Data Analysis of Selected Amaranthus Species from Shirala Tahsil, Dist. Sangli, MS, (India). *International Journal*

for *Innovative Research in Science & Technology*, **7**, 47-55.

- [32] Mwase, W.F., Kachiguma, N., Manduwa, D. and Maliro, M.F.A. (2014) Agromorphological Diversity of Amaranthus Species in Central Malawi. *International Journal of AgriScience*, **4**, 235-241.
- [33] Achigan-Dako, E.G., Sogbohossou, O.E.D. and Maundu, P. (2014) Current Knowledge on Amaranthus Spp.: Research Avenues for Improved Nutritional Value and Yield in Leafy Amaranths in Sub-Saharan Africa. *Euphytica*, **197**, 303-317. <https://doi.org/10.1007/s10681-014-1081-9>
- [34] Molin, W.T. and Nandula, V.K. (2017) Morphological Characterization of *Amaranthus palmeri* x *A. spinosus* Hybrids. *American Journal of Plant Sciences*, **8**, 1499-1510. <https://doi.org/10.4236/ajps.2017.86103>
- [35] Brenner, D.M., Baltensperger, D.D., Kulakow, P.A., Lehmann, J.W., Myers, R.L. and Slabbert, M.M. (2010) Genetic Resources and Breeding of Amaranthus. In: Janick, J., Ed., *Plant Breeding Reviews*. Vol. 19, John Wiley & Sons. Inc., 227-285.
- [36] Andini, R., Yoshida, S., Yoshida, Y. and Ohsawa, R. (2013) Amaranthus Genetic Resources in Indonesia: Morphological and Protein Content Assessment in Comparison with Worldwide Amaranths. *Genetic Resources and Crop Evolution*, **60**, 2115-2128. <https://doi.org/10.1007/s10722-013-9979-y>
- [37] Sagar, K.P., Hanchinamani, C.N., Imamsaheb, S.J., Nishani, S. and Ramanagouda S.H. (2018) Genotypic Correlation Coefficients among Growth Yield and Quality Parameters in Amaranthus Genotypes (*Amaranthus tricolor* L.). *International Journal of Current Microbiology and Applied Sciences*, **7**, 2701-2707. <https://doi.org/10.20546/ijcmas>
- [38] Erum, S., Ambreen F., Naeemullah, M., Masood, S., Qayyum, A. and Rabbani, M.A. (2012) Genetic Divergence in Amaranthus Collected from Pakistan. *The Journal of Animal & Plant Sciences*, **22**, 653-658.
- [39] Das, S. (2016) Amaranthus: A Promising Crop of Future. Springer. <https://doi.org/10.1007/978-981-10-1469-7>
- [40] Yarnia, M., Benam, M.K. and Tabrizi, E.F.M. (2010) Sowing Dates and Density Evaluation of Amaranth (cv. Koniz) as a New Crop. *Journal of Food, Agriculture & Environment*, **8**, 445-448.
- [41] Fageria, N.K., Castro, E.M. and Baligar, V.C. (2004) Response of Upland Rice Genotypes to Soil Acidity. In: Wilson, M.J., He, Z. and Yang, X., Eds., *The Red Soils of China*, Springer, 219-237. https://doi.org/10.1007/978-1-4020-2138-1_14
- [42] Sarker, U. and Oba, S. (2021) Color Attributes, Betacyanin, and Carotenoid Profiles, Bioactive Components, and Radical Quenching Capacity in Selected Amaranthus Gangeticus Leafy Vegetables. *Scientific Reports*, **11**, Article No. 11559. <https://doi.org/10.1038/s41598-021-91157-8>
- [43] Shongwe, V.D., Magongo, B.N., Masarirambi, M.T. and Manyatsi, A.M. (2010) Effects of Irrigation Moisture Regimes on Yield and Quality of Paprika (*Capsicum annum* L.). *Physics and Chemistry of the Earth, Parts A/B/C*, **35**, 717-722. <https://doi.org/10.1016/j.pce.2010.07.038>
- [44] Mekonnen, M.M. and Gerbens-Leenes, W. (2020) The Water Footprint of Global Food Production. *Water*, **12**, Article No. 2696. <https://doi.org/10.3390/w12102696>