

International Journal of Current Microbiology and Applied Sciences ISSN: 2319-7706 Volume 8 Number 02 (2019)

Journal homepage: http://www.ijcmas.com



Original Research Article

https://doi.org/10.20546/ijcmas.2019.802.155

Evaluation of Different Soil Amendments and Germplasm / Varieties against Tomato Bacterial Wilt Caused by *Ralstonia solanacearum*

R.K. Bannihatti*, A.P. Suryawanshi, K.S. Sayyed and V.B. Bhujabal

Department of Plant Pathology, Vasantrao Naik Marathwada Krishi Vidyapeeth, Parbhani-431402, Maharashtra, India

*Corresponding author

ABSTRACT

Keywords

Tomato, Bacterial wilt, Organic amendments, Germplasm/ Varieties, Germination, Pre emergence mortality (PEM), Wilt incidence and Ralstonia solanacearum

Article Info

Accepted:
12 January 2019
Available Online:
10 February 2019

Bacterial wilt caused by Ralstonia solanacearum (Smith) Yabuuchi is one of the most destructive diseases of tomato (Lycopersicum esculentum), causing accountable losses of about 10-90 per cent. Present investigations on the disease (R. solanacearum) were carried out during 2014-15 to fulfill the objectives defined, at the Department of Plant Pathology, College of Agriculture, VNMKV, Parbhani. A total of 10 organic amendments evaluated (pot culture) as pre- sowing soil applications were found effective against R. solanacearum. However, significantly highest seed germination was recorded with vermicompost (76.50 %), followed by karanj cake (71.74 %), neem seed cake (66.75 %) and compost (61.67%); whereas, significantly highest reduction in average incidence (PEM and wilt) was recorded with vermicompost (60.66%), followed by karanj cake (53.74%), neem seed cake (46.70 %) and compost (41.33%). Under artificial epiphytotic conditions (root zone drenching method), all the 14 tomato entries evaluated exhibited different reactions against R. solanacearum. However, six entries (Tom-21, Tom-4, Tom-17, Tom-13, S-22 and PKM-1) were moderately resistant with average wilt incidence in the range of 25.28 to 33.25 per cent; four entries (Tom-15, Tom-11, Tom-27 and Tom-7) were moderately susceptible with average wilt incidence in the range of 47.81 to 52.46 per cent and four entries (Tom-8, Tom-18, Tom-2 and Pusa Ruby) were susceptible with average wilt incidence in the range of 61.13–78.50 per cent.

Introduction

Tomato (Solanum lycopersicum L.) is one of the most widely grown fruit vegetable in the world, with third rank in priority after Potato and Onion in India but ranks second after potato in the world. India ranks second in the area as well as in production of Tomato. Commercially grown throughout the world for fresh fruit, market and processing industries. China is the largest tomato producing country in the world, followed by India and USA (Anonymous, 2014). In India, the area under tomato cultivation was 880 thousand hectare with production of 18227 thousand MT and productivity of 20.7MT/ha (Anonymous, 2013-14). The Maharashtra state is the fourth largest tomato producer in

the India with an area of 50 thousand hectare, production of 1050 thousand MT and productivity 21MT/ha (Anonymous, 2013-14). Other leading tomato producing states are: Andra Pradesh, Karnataka and Orrisa.

In the tropics, tomato production is severely constrained by disease and insect pests. Tomato crop is being affected by many fungal, bacterial, viral and nematode diseases bacterial such as wilt [Ralstonia solanacearum (Smith) Yabuuchi], bacterial leaf spot (Xanthomonas compestris pv. vesitocoria), bacterial canker (Clavibacterm ichiganensi spv. michiganensi), early blight (Alterneria solani), powdery mildew (Leveillula taurica)Tomato mosaic virus, Tomato leaf curl virus and Tomato spotted wilt (viruses) and root knot nematode (Meloidogyne incognata).

Among these diseases, bacterial wilt caused (Smith.) by Ralstonia solanacearum Yabuuchi (formerly Pseudomonas solanacearum) is one of the most economically important and devastating disease of tomato crop. The disease was first reported from Asia and South America (Smith, 1880). This disease is of common occurrence whenever solanaceous crops viz tomato, brinjal, potato and chillietc are grown and is more severe under weather conditions of high temperature and high humidity, congenial for disease development (Sunder et al., 2011).

In India bacterial wilt of tomato was first reported in Solan area of Himachal Pradesh(Gupta *et al.*,1998) *R.solanacearum* (Smith) is a serious soil borne pathogen of solanaceous vegetable crops grown during summer, rainy and winter seasons. Tomato (*Lycopersicon esculentum*) is one of the important solanaceous vegetables, which suffers badly due to *R solanacearum*, wherever high temperature (28 to 36°C) and high moisture (50 to 100 %) prevails (Sharma

et al., 2009). In India about 10 to 100% incidence of tomato bacterial wilt during the summer were reported (Kishun, 1985). R. solanacearum is a globaly dispersed and heterogeneous bacterial pathogen, with socioeconomic impacts (Yabuuchi et al., 1995).

Materials and Methods

Bioefficacy of organic amendments

A total of 9 organic amendments were evaluated against *R solanacearum* by sick soil method in pot culture, under screen house conditions. The test amendments were applied as presowing treatment (protective). Except vermicompost, all the test amendments were crushed physically to coarse form and used for soil application. The earthen pots (30 cm dia.) disinfected with 5 per cent solution of Copper sulphate were filled with autoclaved potting mixture of soil: sand: FYM (2:1:1).

The mass multiplied (48 hr old nutrient broth culture: 2 x10⁸ cfu/ml) of *R* solanacearum was drenched (@ 50 ml/ kg potting mixture) evenly to the potting mixture in pots, these pots were incubated for 96 hrs in screen house to proliferate the bacterium and make the soil / potting mixture sick. The coarse ground test amendments were applied (@ 50 g / kg mixture) in the earthen pots containing test bacterium sick soil/ potting mixture, mixed thoroughly, watered regularly and maintained in screen house. After 72 hrs of amendments application, surface sterilized (0.1 % HgCl₂) healthy seed of tomato Cv. Pusa Ruby were sown (20 seeds/pot), watered regularly and maintained in the screen house. Three pots / treatment / replication were maintained. The earthen pots containing R solanacearum sick soil and sown with surface sterilized healthy seed of tomato cv. Pusa Ruby, without amendment were maintained as untreated control.

Experimental details

Design : CRD
Replications : Three
Treatments : Ten

Variety : Pusa Ruby

Treatment details

T₁: Compost

T₆: Groundnut cake

T₂:Poultry manure T₇: Sunflower cake

T₃:Vermicompost T₈:Cotton seed cake

T₄:Goatmanure T₉: Neem seed cake

 T_5 :Karanja cake T_{10} : Control (untreated)

Observations on seed germination and preemergence mortality (PESR) were recorded at seven days after sowing and that of wilting were recorded at 30th and 45 DAS. The per cent seed germination, pre-emergence mortality (PEM) and wilting were calculated by following formulae:

Germination (%) =

No. of seeds germinated x 100 Total no. of seeds sown

$$\label{eq:wilted} Wilting (\%) = \frac{\text{No. of seedlings died/wilted}}{\text{Total no. of seedlings}} \times 100$$

Where,

C= Per cent mortality /wilting in treatment pots

T = Per cent mortality / wilting control pots

Screening of tomato entries

Fifteen days old seedlings of the 14 test entries were transplanted (5 seedlings/pot/entry) in the earthen pots (30 cm dia.) filled with steam sterilized potting mixture of soil: sand: FYM (2:1:1), maintained in screen house and watered regularly.

Two pots/ entry/replication were maintained. One week after transplanting these potted seedlings were inoculated by drenching at root zone with 48hr old pure culture suspensions of R.soalancearum (2×10 8 cfu/ml) and maintained by watering frequently, under screen house.

Observations on bacterial wilt incidence were recorded applying 0-5 grade disease rating scale (Winsted and Kelman, 1952) at 30 and 45 days after inoculation of pathogen. The data was averaged and percent bacterial wilt disease incidence was calculated by following formula.

Percent disease incidence (PDI) =

Number of plants showing wilts symptoms
-----X 100
Total number of plants

Disease rating scale

Grade	% Incidence	Disease Reactions				
0	Highly resistant (HR)	Plants did not show any wilt symptom				
1	Resistant (R)	1 - 20 % plants wilted				
2	Moderately resistant (MR)	21- 40 % plants wilted				
3	Moderately susceptible (MS)	41- 60 % plants wilted				
4	Susceptible (S)	61- 80% plants wilted				
5	Highly susceptible (HS)	More than 80% plant wilted				

Experimental details

Design : C.R.D.
Replications : Three
Treatments : Fourteen

Tr. No	Treatments	Tr. No	Treatments
T ₁	Tom-5	T ₈	Tom 18
T ₂	Tom 11	T ₉	Tom 2
T ₃	Tom 21	T_{10}	Tom 17
T ₄	Tom 8	T ₁₁	Tom 13
T ₅	Tom 27	T ₁₂	S-22
T ₆	Tom 7	T ₁₃	PKM-1
T ₇	Tom 4	T ₁₄	Pusa Ruby

Statistical analysis

The data obtained in all the experiments was statistical analyzed. The percentage values were transformed into arcsine values.

The standard error (S.E.) and critical difference (C.D.) at level P = 0.01 were worked out and results obtained were compared statistically. All the statistical analysis was done using VNMKV-STAT statistical programmer at Central Computer Laboratory, Vasantrao Naik Marathwada Krishi Vidyapeeth, Parbhani.

Results and Discussion

In vitro bioefficacy of organic amendments

A total of 9 amendments were evaluated as pre-sowing soil application to assess their efficacy against *R solanacearum*, employing sick soil technique and sowing susceptible tomato cv. Pusa Ruby in pot culture under glass house conditions and the results obtained on seed germination, pre-emergence mortality and wilting are presented in the Table 1 and Fig.1.

Seed germination

Results (Table 1) revealed that all the test amendments significantly improved the per cent seed germination, over untreated control and it was ranged from 40 to 76.50 per cent, as against 35 per cent in untreatedcontrol. However, vermicompost and karanj cake were found most effective with significantly highest seed germination of 76.50 and 71.74 per cent, respectively and both were at par. These were followed by the amendments viz.,neem seed cake (66.75 %),compost (61.67 %),sunflower cake (56.72%), Goat manure (51.55) and Cotton seed cake (48.25%) respectively and later two were at par. The amendments viz., poultry manureand groundnut cakes were found comparatively least effective with minimum germination of 40.0 and 43.29 per cent, respectively.

Pre emergence mortality

Results (Table 1 and Fig) revealed that all the test amendmentssignificantly influenced the pre-emergence mortality (PEM) and it was ranged from 23.33 to 60.00 per cent, as against 65.00 per cent in untreated control. However, vermicompostwas found most with significantly least effective preemergence mortality (23.33 %), followed bykaranj cake (28.40%), neem seed cake (33.25%), compost (38.33%), sunflower cake (43.41%), goat manure (48.29%) and Cotton seed cake (51.67%). Rest of the amendments recorded pre emergence mortality 56.67 to 60.00 per cent respectively as against 65.00 per cent in control.

Wilt incidence

Percent wilting recorded with all the test amendments was from 30.55 to 65.27 per cent, as against 71.42 per cent in untreated control. However, vermicompost was found most effective with significantly least wilting

per cent (30.55 %), followed by amendments karanj cake (34.92) and neem seed cake(36.65 %) both were at par; compost (40.59%), sunflower cake (50.25%), goat manure (51.51%) and cotton seed cake (55.55%). Rest of the amendments groundnut cake and poultry manure were least effective and recorded comparatively maximum wilting per cent in the range of 61.57 to 65.27 per cent, respectively.

Average (PEM and Wilt) recorded in all the test amendments were ranged from 26.94 to 62.63 per cent, as against 68.21 per cent in untreated control.

However, comparatively minimum average incidence was recorded with vermicompost (26.94%), followed by karanj cake (31.66%), neem seed cake (35.00%), compost (39.46%), sunflower cake (46.83%), goat manure (49.90%) and cotton seed cake (53.61 %). Rest of the amendments groundnut cake and poultry manure were recorded average incidence in the range of 59.12 to 62.63 per cent, respectively.

Reduction in mortality and wilt incidence

All test amendments recorded significant reduction in pre -emergence mortality (PEM) over untreated control. Reduction in pre emergence mortality recorded was ranged from 7.67 to 64.10 per cent.

However, significantly highest reduction in pre emergence mortality (PEM) was recorded in vermicompost (64.10%), followed karanj cake (56.40%), neem seed cake (48.72%), compost (39.42%), sunflower cake (33.33%), goat manure (25.63%) and cotton seed cake %). Rests of the amendments (21.81)groundnut cake and poultry manure were recorded least reduction pre emergence mortality in the range of 12.82 to 7.90 per amendments cent. All test recorded

significant reduction in percent wilting over control. Reduction in percent wilting recorded was ranged from 8.65 to 57.22 per cent. However, significantly highest per cent wilting reduction in recorded vermicompost (57.22 %), followed by karanj cake (53.73%), neem seed cake (45.20 %), compost (43.15%), sunflower cake (29.64%), goat manure (28.77%), and cotton seed cake(23.90%). Rests of the amendments groundnut cake and poultry manure were found comparatively less effective with reduction percent wilt recorded was from the range of 20.98 to 8.65 per cent.

Average reduction in the incidence (PEM and Wilt) recorded in the test amendments were ranged from 8.17 to 60.66 per cent over untreated control.

However, significantly highest reduction was recorded with vermicompost (60.66%), followed by karanj cake (53.74%), neem seed cake (46.7 %), compost (41.33%), sunflower cake (29.98%), goat manure (27.20%) and cotton seed cake (22.87%).

Rests of the amendments groundnut cake and poultry manure were found comparatively less effective with reduction in average incidence was range from 16.90 to 8.67 per cent.

Results of the present study obtained on efficacy of organic amendments *viz.*, vermicompost, karanj cake, neem seed cake, compost, sunflower seed cake, goat manure, cotton seed cake, groundnut cake and poultry manure against *R. solanacearum* are in conformity with those reported earlier by several workers (Sharma and Kumar, 2000; Sharma and Kumar, 2004; Islam and Toyota, 2004; Bose *et al.*, 2004; Messiha *et al.*, 2007; Sharma and Kumar, 2009; Ghosh *et al.*, 2009., Yadessa *et al.*, 2010; Reddy *et al.*, 2012 and Djeugap *et al.*, 2014)

Table.1 Efficacy of organic amendments against *R. solanacearum*

Tr. No	Treatments	Germin ation* (%)	Incidence* (%)		Av.	Reduction over control (%)		Av. (%)
			PEM	Wilt	(%)	PEM	Wilt	
T ₁	Compost	61.67 (51.75)	38.33 (38.25)	40.59 (39.58)	39.46 (38.92)	39.52 (38.95)	43.15 (41.06)	41.33 (40.01)
T ₂	Poultry manure	40.00 (39.23)	60.00 (50.77)	65.27 (53.89)	62.63 (52.32)	7.69 (16.10)	8.65 (17.10)	8.17 (16.61)
T ₃	Vermicompost	76.50 (61.00)	23.33 (28.88)	30.55 (33.55)	26.94 (31.27)	64.10 (53.19)	57.22 (49.15)	60.66 (51.15)
T ₄	Goat manure	51.55 (45.89)	48.29 (4.024)	51.51 (45.87)	49.90 (44.94)	25.63 (30.42)	28.77 (32.44)	27.20 (31.44)
T ₅	Karanj cake	71.74 (47.89)	28.40 (32.20)	34.92 (36.22)	31.66 (34.24)	56.40 (48.68)	51.09 (45.62)	53.74 (47.14)
T ₆	Groundnut cake	43.29 (41.14)	56.67 (48.83)	61.57 (51.69)	59.12 (50.25)	12.82 (20.98)	20.98 (27.26)	16.90 (24.27)
T ₇	Sunflower cake	56.72 (48.86)	43.41 (41.21)	50.25 (45.14)	46.83 (43.18)	33.33 (35.26)	29.64 (32.99)	29.98 (32.20)
T ₈	Cottonseed cake	48.25 (44.00)	51.67 (45.96)	55.55 (48.19)	53.61 (47.07)	21.84 (27.86)	23.90 (29.27)	22.87 (28.27)
T ₉	Neem seed cake	66.75 (54.79)	33.25 (35.21)	36.75 (37.32)	35.00 (36.27)	48.72 (44.27)	45.20 (42.25)	46.7 (43.11)
T ₁₀	Control (Untreated)	35.00 (36.27)	65.00 (53.73)	71.42 (57.68)	68.21 (55.68)			
	SE ±	1.75	1.75	2.76	2.25	2.15	2.40	2.27
	CD (P= 0.01 %)	5.21	5.21	8.24	6.75	6.41	7.14	6.82

^{*}Means of three replications, Figures in parenthesis are arcsine transformed value, Av = Average, PEM = Pre emergence mortality

Table.2 Reactions of tomato genotypes, germplasm lines, cultivars and varieties against *R. solanacearum*(pot culture)

Tr. No	Treatment /Entries	Disease incidence (%)		Average	Varietal	
		30 DAT	45DAT	incidence	reactions	
T1	Tom -15	33.33	65.33	49.33	MS	
T2	Tom -11	67.72	37.17	52.46	MS	
T3	Tom -21	24.12	30.00	27.06	MR	
T4	Tom -8	63.50	67.20	65.35	S	
T5	Tom -27	67.17	33.55	50.33	MS	
T6	Tom -7	26.50	69.13	47.81	MS	
T7	Tom -4	29.33	31.83	30.58	MR	
T8	Tom -18	59.43	62.83	61.13	S	
T9	Tom -2	65.00	67.83	66.41	S	
T10	Tom -17	22.23	28.33	25.28	MR	
T11	Tom -13	22.13	33.33	27.73	MR	
T12	S-22	27.50	32.00	29.75	MR	
T13	PKM-1	31.37	35.13	33.25	MR	
T14	Pusa ruby	79.00	78.00	78.5	S	
	SE ±	0.75	0.40	0.57		
	CD	2.24	1.17	1.71		

DAT- Days after transplanting, S- Susceptible, MS – Moderately susceptible and MR- Moderately resistant

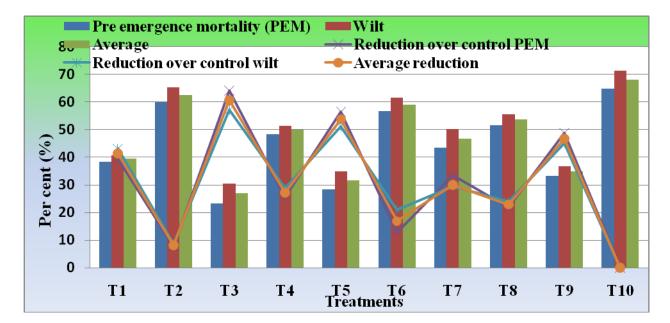


Fig.1 In vitro efficacy various organic amendments against R. solanacearum

Evaluation of tomato test entries against *R. solanacearum*

Results (Table 2) revealed the under artificial epiphytotics and controlled conditions of the screen house, all 14 tomato entries exhibited different reactions against R solanacearum. However, six test entries Tom-21, Tom-13, Tom-17, S-22 and PKM -1 were found moderately resistant with average bacterial wilt percent disease incidence in the range from 25.28 to 50.33per cent; while four test entries Tom-7, Tom-15, Tom-11 and Tom-27 were found moderately susceptible bacterial wilt with average percent disease incidence in the range from 47.81 to 52.46 per cent; whereas, four test entries Tom-8, Tom-18 Tom-2 and Pusa Ruby found susceptible with average percent wilt incidence in the range 61.13 to 78.5 percent. Pusa Ruby was found susceptible to the disease bacterial wilt with maximum per cent disease incidence (64.36 %). None of the entry was found highly resistant or immune to the bacterial wilt of tomato disease. These results obtained on varied reactions of the tomato test entires against R solanacearum are on the same line

as to that of reported earlier by several workers (Khan *et al.*, 1974; Kapoor *et al.*, 1991; Singh and Sood, 2003; Sudheendra *et al.*, 2003; Biswas and Singh 2007; Matsunga *et al.*, 2011; Myint, 2011; Artal *et al.*, 2012; Dutta and Rahman 2012; Dutta *et al.*, 2013 and Pawaskar *et al.*, 2014).

References

Anonymous, 2013-14. Indian Horticultural Databse.www.indiastat.com

Artal, R.B., Gopalakrishnan, C. and B. Thippeswamy. 2012. An efficient inoculation method to screen tomato, brinjal and chilli entries for bacterial wilt resistance.Pest Management Horti Ecosystems. Vol. 18(1):70-73.

Biswas, S. and Singh, N. P. 2007. Effect of host genotype and cultural practice for the management of bacterial wilt in brinjal (*Solanaummelongena*). Indian phytopath. 60 (4): 438 – 441.

Bose, T.K., Kabir, J., Maity, T.K., Parthasarrthy, V. A. and Som, M.G. 2004. Vegetables Crops (vol. I edt), NayaPrakash, Kolkatta: 987.

- Djeugap, J.F., Eko, D., Julienne, J., Columbus, T.N. and Fonte, A.D. 2014. Effect of organic amendments and fungicide application on potato late blight, bacterial wilt and yield in Cameroon. Int. J. Agro. Agric. Res. 5 (4): 12-19
- Dutta, P. and Rahman, B. 2012. Varietal screening of tomato against bacterial wilt disease under subtropical humid climate of Tripura. Inter. J. Farm Sci. 2(2): 40-43.
- Dutta, R., Satish Chandra.and Ngachan, S.V. 2013.Evaluation of tomato genotypes against bacterial wilt (*Ralstonia solanacearum*) under mid-hill conditions.Indian Phytopath. 66 (1): 96-97
- Ghosh.P.P. and Mandal, N. C. 2009. Some disease management practices for bacterial wilt of potato. J. Pl. Prot. Sci. 1(1): 51-54.
- Gupta, S.K., Dohroo, N.P and Shyam, K.R. 1998. Occurance of bacterial wilt of tomato in Himachal Pradesh. Pl.Dis.Res.13: 174.
- Islam T.M.D. and Toyota K. 2004. Suppression of bacterial wilt of Ralstonia tomato caused by by incorporation of solanacearum soil composts and possible in mechanism.Microbes Environment. 19:53-60.
- Kapoor, A.S., Sugha, S.K. and Dhambir, Dhambir Singh. 1991. Partial resistance to bacterial wilt in tomato. Indian Phytopath. 44: 224-245.
- Khan, A.N.A., PatilKulakarni, B.G and Hegde, R.K. 1974.Partial resistance to bacterial resistance to bacterial wilt in brinjal and tomato.Curr.Res. 10: 30-31.
- Kishun, R.1981. Studies on bacterial wilt of solanaceous crops. Ann. Sci. Rept. IIHR, Bangalore. 18-22.
- Kishun, R. 1985. Effect of bacteria wilt on yield of tomato.Indian phytopathol.38:

- 606.
- Matsunga, H., Saito, T. and Saito, A. 2011. Evaluation of resistance to bacterial wilt and Phytophthora blight in *Capsicum* resources collected in Myanmar. J. Japan. Soc. Hort. Sci. 80 (4): 426-433.
- Messiha, N.A.S., Ariena H. C., van Bruggen., Anne D. van Diepeningen., Oscar J. deVos., J. Termorshuizen., N. N.A., Tjou-Tam-Sin.and Janse, J. D.2007. Potato brown rot incidence and severity under different management and amendment regimes in different soil types. Eur. J. Plant. Patho. 119:367–381
- Myint, W.W. 2011. Study on the response of three tomato cultivars tested with *Ralstonia solanacearum*causing bacterial wilt of tomato. Yangon University of DistanceEdun. Res. J. 3 (1):125-133.
- Pawaskar, Kadam, J.R., Navathe, S.and Kadam J.S. 2014. Response of chilli varieties and genotypes to bacterial wilt caused By *Ralstonia solanacearum* and its Management. Indian J. Sci. 11(29): 66-72.
- Reddy, S.A., Joseph, D., Bagyaraj.and Kale, R.D. 2012. Vermicompost as a biocontrol agent in suppression of two soil-borne plant pathogens in the field. Acta Biological Indica: 137-142.
- Sharma, J.P. and Kumar. S. 2000. Management of *Ralstonia* wilt through soil disinfectants, mulch, lime and cakes in tomato (*Lycopersiciescculentum*). Indian J. Agrc. Sci. 70: 17-19.
- Sharma, J. P. and Kumar, S. 2009a. Linear reduction of propagules of *Ralstonia solanacearum*in soil by cake and chemicals.Indian Phytopath, 62(1): 49-53.
- Sharma, J.P. and Kumar, S. 2009b. Management of *Ralstonia*wilts of tomato through microbes, plant extracts and combination of cake and chemicals,

- Indian phytopath. 62 (4): 417-423.
- Singh, Y. and Sood, B. 2003. Screening of sweet pepper germplasm for resistance to bacterial wilt (R.solanacearum). Eggplant Newsl. 22: 117-120.
- Smith, E.F. (1880). A bacterial diseases of the tomato, potato, eggplant and Irish potato (*Bacillus solanacearum*nov sp.) U. S. Dept. Agric. Div. Veg. Physiol, Path. 12: 1-28.
- Sudheendra, A., Ashtaputre, A.M. and Rao, M.S.L. (2003). Evaluation of brinjal varieties against bacterial wilt. In: Procedings of recent development in the diagnosis and management of plant diseases for meeting global challenges. U.A.S.

- Dharwad. 18-20, 2003.
- Winstead.N.N. and Kelmen, A. (1952).Inoculation techniques for evaluating resistances to *Pseudomonas* solanacerum.Phytopatho.42:628 -634.
- YabuuchiE., Kosako Y., Yano I., Hota H.and Nishiuchi Y. 1995. Transfer of two Burkholderiaand an Alcaligenes species to Ralstoniagen. nov.Ralstonia solanacearum (Smith, 1986) Microbiol and Immnl.39: 897-904.
- Yadessa, G.B., Bruggen, A. H. C. and Ocho, F.L. 2010. Effects of different soil amendments on bacterial wilt caused by *Ralstonia solanacearum and* on the yield of tomato. J. Pl. Pathol. 92(2): 439-450

How to cite this article:

Bannihatti, R.K., A.P. Suryawanshi, K.S. Sayyed and Bhujabal, V.B. 2019. Evaluation of Different Soil Amendments and Germplasm / Varieties against Tomato Bacterial Wilt Caused by *Ralstonia solanacearum*. *Int.J.Curr.Microbiol.App.Sci.* 8(02): 1331-1339.

doi: https://doi.org/10.20546/ijcmas.2019.802.155